



THE ROLE OF HERBAL TO IMPROVE BROILER PERFORMANCE: A SUBJECT REVIEW

Sawsan Abdul Faraj Mohammed
Collage of Nursing, University of Kirkuk, Iraq
sawsanabd@uokirkuk.edu.iq

Abstract

This review was conducted to explain the efficacy of some herbals as a growth and performance promoter in broiler. The importance and use of herbal remedies has been reviewed systematically. Garlic can be utilized to successfully increase broiler growth, Aloe vera has been shown to be beneficial in the treatment of wounds and the treatment of diabetes. Fenugreek has the ability to provide ruminant animals with high-quality feed. Nettle leaves were found to boost body weight increase and feed conversion efficiency for broilers. The effects of Thyme extracts or essential oils on a range of meat varieties, including pig, beef, and lamb, have been thoroughly examined and reported after their direct addition. The weight gain of Cobb broilers is dramatically increased by 16.97% when *O. sanctum* aqueous extracts is added to water compared to the controls.

Keywords: herbal; Garlic; Aloe vera; Fenugreek; Nettle; broiler.

Introduction

Antibiotics have proven to be the most cost-effective method of maintaining feed efficiency and health in monogastrics such as chicken [1]. However, because human activities are continuously evolving or being influenced, things are certain to change throughout time as a result of their use. As a result, the increased and purposeful interest in utilized of herbs and other plant products in modern poultry production is an intriguing development. This is consistent with millennial farming's 'clean' production practices. The hunt for synthetic medication alternatives has intensified as a result of this [2]. Plant extracts can help with both your health and your overall performance [3-8]. Increased endogenous proteolytic enzyme secretion, stimulation of appetite and nutrient intake, and activation of the immune system, and the antiviral effects, with antioxidant properties, and the antihelminthic effects could all have a good impact on animal health and performance. Fenugreek seeds have been discovered to be high in protein, fat, total carbs, and minerals like calcium ion, the phosphorus, and the iron, with zinc, and magnesium. Fenugreek also helps the gastrointestinal tract with its accessories [9-10]. In hens, it was discovered that





phytogenic feed additions increased weight gain, feed conversion rate, and viability [11]. Ewuola and Egbunike [12] reported that Some herbal medicines are primarily utilized as nutritional supplement or medicines, resulting in a cascade of the physiological reactions that may result in serum metabolite changes. This could be due to toxic substances in the plant causing a decrease or increase in serum metabolites. It might act as non-toxic chemical components that keep chicken qualities inside the expected reference ranges [13]. This review was conducted to explain the efficacy of some herbals as a growth and performance promoter in broiler.

Garlic

Garlic (*Allium sativum*) contains a variety of bioactive compounds that are useful to the body [14]. According to Ziarlarimi et al [15], garlic powder (GP) contains several organosulfur compounds like as allicin compound, alliin, the ajoene compound, and the S-allylcysteine compound, demonstrating that adding garlic to feed like a healthy plant-derived feed supplement can effectively boost broiler growth [16]. Since antiquity, garlic has also been used as a medicinal herb in the management and treating a wide variety of heart, gastrointestinal, and metabolic disorders, including diabetes, dementia disorders, tumors, and plaques of blood vessels [17]. Garlic has antibacterial, antioxidant, antithrombotic, antiplatelet aggregator, and antihypertensive activities in broilers, according to several studies [18-19].

Aloe vera (*Aloe barbadensis*)

It is an Asphodelaceae (Liliaceae) shrubby, or absorescent, perennial, tropical plant with a pea-green colour. It has large, fleshy triangular leaves with spikes along the edges [20]. Aloe vera has been shown to be beneficial in the treatment of wounds [21] and to hasten wound healing [22]. Aloe vera can also be used topically to treat genital herpes and psoriasis [21]. Because of the inclusion of chemicals such as mannans, anthraquinones, and lectins, Aloe vera derivatives may be useful for treating human diabetes and high blood lipid levels [23]. Oral aloe vera gel may improve symptoms and inflammation in ulcerative colitis patients, according to preliminary research [24]. Aloe vera extracts have been utilized as an immunostimulant to help cats and canines fight cancer [25]. Aloe vera extracts might have antibacterial and antifungal actions that could help treat skin disorders including boils and skin cysts and stop the development of the fungi that cause tinea [26].





Fenugreek

Fenugreek is single-cut annual legume forage that has been farmed in Southern Europe countries, the Asia countries, and the countries of Northern Africa for centuries [27]. It was first introduced to Canada in the early 1990s. Seed production potential [28], fodder yield and nutrient content [29-30], and medicinal value [31] of fenugreek planted on the Canadian prairies has all been studied. These researches have highlighted fenugreek's immense potential as an alternative feed crop for temperate climates, such as the Western Canadian prairie. Apart from crop cultivation, fenugreek's annual nature may allow growers to use it as a break crop in farming systems [32]. Fenugreek has the ability to provide ruminant animals with high-quality feed. Indeed, in the Mediterranean region, fenugreek was traditionally used as a feed crop [27], and the species name 'foenum graecum' means 'greek hay' [33]. There are few studies that have used fenugreek fodder to feed cattle. Beef steers fed fenugreek silage dietary supplementation with barley grain had equivalent diabetes complications intake, daily average growth, and feed performance in comparison to steers fed alfalfa silage supplemented with seeds [29].

Nettle

Urticadioica, sometimes known as nettle, is a perennial plant that has medicinal properties. It has been shown that nettle can improve the immune system of fish [34], protect rabbit and poultry from endo- and ectoparasites [35], and influence blood lipid profile [36]. Nettle leaves contain carotenoids, vital fatty acids, various vitamins, different types of minerals, phytosterol compounds, glycosides, and proteins, which have the potential to be used in the feed, medicinal, and cosmetic industries [37]. As a feeding approach for broilers, nettle leaves can be substituted for soybean meal up to 9% of the time [38]. Despite the fact that nettle leaves have a high iron content and are a rich source of calcium and vitamin A, they can be used to treat anemia [39]. By the way, nettle extracts have been shown to have antimicrobial properties [40]. The chemicals utilized in nettle extraction, on the other hand, have a significant antibacterial effect on the final product [41]. At the age of 21-42, nettle leaves were found to boost weight gain and nutrient conversion rate [42]. Meanwhile, it has been found that nettle leaves did not improve the oxidative stress in the broiler's body [43]. This suggests that nettle could be a useful additive in poultry feeding for growth enhancement. The nettle leaf extracts (NLE) is a nettle product. While NLE has been proven to be effective against fungi [44], more research into its effects on microorganisms is needed [45]. Antibiotics are no longer used as a growth booster in





poultry feeds in several countries due to antibiotic-induced microbial resistance. The extraction process improves the quality of NLE significantly [46].

Thyme

The effects of the herbal extracts or essential oils on a range of meat varieties, including pig [47], beef [48], and lamb [49], have been thoroughly examined and reported after their direct addition. In addition, various studies [50-53] have shown that diets including rosemary compounds and the oregano can reduce lipid oxidation and microbiological counts in a variety of meats. The usefulness of thyme in the diet, like is the case with thyme, is less well understood. Thyme essential oil (EO) includes about 60 components, the majority of which having therapeutic effects such as antibacterial, carminative, antioxidant, and antimicrobial. The thymol (68.1%) and carvacrol compound (3.5%) are the most abundant and active chemicals in thyme EO, along with the (11.2%) monoterpene hydrocarbons p-cymene and (4.8%) c-terpinene compound [54], which have antioxidant and antibacterial effects. These chemicals' antibacterial activities are linked to their lipophilic nature, which leads to their accumulation in membrane surface, where they take part in later membrane-related processes such energy exhaustion. Furthermore, Polyphenolic redox properties enable them to act as reductants, donors of hydrogen, metal chelators and the singlet oxygen quenchers [55]. In general, dietary supplementation has proven to be a straightforward and practical method of introducing antioxidant enzymes into phospholipid, where they can effectively block oxidative processes in situ [56].

Tulsi (*Ocimum sanctum*)

Ocimum sanctum, a member of the Lamiaceaceae family, is well known for its medicinal properties. Tulsi and Holy basil are other names for it. It thrives in the tropical regions and warm climates. It is widely dispersed and grown in India. It is an erect, lateral branches, fragrant plant that grows to a height of 30-60cm when mature [57]. The leaves of *O. sanctum* can be fed as a powder form mixed with feed or as an extract blended in water. According to Hasan et al. [58], adding *O. sanctum* aqueous extracts to water causes a substantial 16.97% rise in the body weight of cob broilers compared to the control group. Broilers dietary supplementation with *O. sanctum* extract of leaves 1ml/liter in tap water showed a higher body weight compared to the control group, according to Biswas et al. [59]. According to Alom et al. [60], supplementing broiler drinking water with Tulsi leaf 2ml/liter leads to noticeably higher body weight. Similar experiments have demonstrated that feeding dry powdered *O. sanctum* leaves to broilers caused a considerable rise in their body





weight. According to Singh et al. [61], broiler chicks fed *O. sanctum* leaf powder at a 1 percent level in feed experienced a substantial rise in the muscular weight of their legs, thighs, and breasts, resulting in a higher weight gain than control chicks. By giving immunely suppressed broiler birds 3g/Kg of *O. sanctum* dry leaves for two weeks between the fifth and sixth week, Mode et al. [62] showed that the birds were protected from immunological suppression and also met the needed target by significantly increasing their weight gain. In comparison to the control group, 15-day treatment of poultry with 200 mg of leaves extract per bird showed the greatest weight gain, according to Gupta & Charan [63]. At this dose level, the chickens did not exhibit any gross or clinically harmful hematological, biochemical, histopathological, or biochemical side effects. The effect of tulsi leaf powder given at 5g/kg feed on weight growth associated with colibacillosis was observed by Kumari [64]. Body weight increase was seen to be considerably higher in the *O. sanctum* leaf treated groups (both infected and uninfected) than in the corresponding non-*O. sanctum* leaf treated control groups.

References

1. Looft T, Johnson TA, Allen HK, et al. In-feed antibiotic effects on the swine intestinal microbiome. *Proc Natl Acad Sci USA*. 2012;109(5):1691–1696.
2. Adegbeye MJ, Elghandour MMY, Faniyi TO, et al. Antimicrobial and antihelminthic impacts of black cumin, pawpaw and mustard seeds in livestock production and health. *Agroforest Syst*. 2018.
3. Janssen, A.M. (1989) Antimicrobial Activities of Essential Oils: A Pharmacognostical Study. Dissertation, Rijks Universiteit, Leiden.
4. Horton, G.M.J., Fennell, M.J. and Prasad, B.M. (1991) Effect of Dietary Garlic (*Allium sativum*) on Performance, Carcass Composition and Blood Chemistry Changes in Broiler Chickens. *Canadian Journal of Animal Science*, 71, 939-942.
5. Bakhiet, A.O. and Adam, S.E.I. (1995) Therapeutic Utility, Constituents and Toxicity of Some Medicinal Plants: A Review. *Veterinary Human Toxicology*, 37, 255-258.
6. Skrabka-Blotnicka, T., Rosin'ski, A., Przysie-Zzna, E., Woloszyn, J. and Elminowska-Wenda, G. (1997) Effect of Dietary Formulation Supplemented with Herbal Mixture on Goose Breast Muscle Quality. Report I. The Effect on the Chemical Composition. *Archives für Geflügelkunde*, 61, 135-138.
7. Gill, A., Delaquis, P. Russo, P. and Holley, R. (2002) Evaluation of Antilisterial Action of Cilantro Oil on Vacuum Packed Ham. *International Journal of Food Microbiology*, 73, 83-92.





8. Manzanilla, E.G., Baucells, F., Kamel, C., Morales, J., Perez, J.F. and Gasa, J. (2001) Effects of Plant Extracts on the Performance and Lower Gut Microflora of Early Weaned Piglets. *Journal of Animal Science*, 1, 473. (Abstract).
9. Gupta, K., Thakral, S., Arora, K. and Chowdhary, M. (1996) Structural Carbohydrate and Mineral Seeds. *Indian Coca Arecenut and Species Journal*, 20, 120.
10. Raju, J., Patlolla, J.M.R., Swamy, M.V. and Rao, C.V. (2004) Diosgenin, a Steroid Saponins of *Trigonella foenum graecum* (Feungreek), Inhibits Azoxymethane-Induced Aberrant Crypt Foci Formation in F344 Rats and Induces Apoptosis in HT-29 Human Colon Cancer Cells. *Cancer Epidemiology, Biomarkers & Prevention*, 13, 1392-1398.
11. Javandel F, Navidshad B, Seifdavati J, Pourrahimi GH, Baniyaghoub S. The favorite dosage of garlic meal as a feed additive in broiler chickens ratios. *Pak J Biol Sci* 2008;11:1746-9.
12. Ewuola, E. O. and Egbunike, G. N. (2008). Haematological and serum biochemical growing rabbit bucks fed dietary fumonisin B1. *African Journal of Biotechnology*, 7(23): 4304 – 4309.
13. Simaraks, S., Chinrasri, O. and Aengwanich, W. (2004). Hematological, electrolyte and serum biochemical values of the Thai indigenous chickens (*Gallus domesticus*) in northeastern, Thailand. *Songklanakarinn Journal of Science and Technology*, 26(3): 425 – 430.
14. Khan RU, Nikousefat Z, Tufarelli V, Naz S, Javdani M, Laudadio V. Garlic (*Allium sativum*) supplementation in poultry diets: effect on production and physiology. *Worlds Poult Sci J* 2012; 68:417-24.
15. Ziarlarimi A, Irani M, Gharahveysi S, Rahmani Z. Investigation of antibacterial effects of garlic (*Allium sativum*), mint (*Menthe spp.*) and onion (*Allium cepa*) herbal extracts on *Escherichia coli* isolated from broiler chickens. *Afr J Biotechnol* 2011;10:10320-2.
16. Teshika JD, Zakariyyah AM, Zaynab T, et al. Traditional and modern uses of onion bulb (*Allium cepa* L.): a systematic review. *Crit Rev Food Sci Nutr* 2019;59(Suppl 1):S39-70.
17. Kim YJ, Jin SK, Yang HS. Effect of dietary garlic bulb and husk on the physicochemical properties of chicken meat. *Poult Sci* 2009;88:398-405.
18. Stanaćev V, Glamočić D, Milošević N, Puvača N, Stanaćev V, Plavša N. Effect of garlic (*Allium sativum* L.) in fattening chicks nutrition. *Afr J Agric Res* 2011;6:943-8



19. Ur Rahman S, Khan S, Chand N, Sadique U, Khan RU. In vivo effects of *Allium cepa* L. on the selected gut microflora and intestinal histomorphology in broiler. *Acta Histochem* 2017; 119:446-50.
20. Foster, S. (1999) *Aloe*. *Herbs for Health.*, 59-60.
21. Vogler, B.K. and Ernst, E. (1999) *Aloe vera*: a systematic review of its clinical effectiveness. *British Journal of General Practice.*, Oct;49:(447):823-8.
22. Hegggers, J.P., Elzaim, H. and Garfield, R. (1997) "Effect of the combination of *Aloe vera*, nitroglycerin, and L-NAME on wound healing in the rat excisional model". *Journal of alternative and complementary medicine.*, 3 (2): 149–53.
23. Boudreau, M.D. and Beland, F.A. (2006) An evaluation of the biological and toxicological properties of *Aloe barbadensis* (miller), *Aloe vera*. *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev.*, 24(1): 103-54.
24. Langmead, L., Feakins, R.M. and Goldthorpe, S. (2004) Randomized, double-blind, placebo-controlled trial of oral *aloe vera* gel for active ulcerative colitis. *Alimentary Pharmacology & Therapeutics.*, 19 (7): 739–47
25. King, G.K., Yates, K.M. and Greenlee, P.G. (1995) The effect of *Acemannan* Immunostimulant in combination with surgery and radiation therapy on spontaneous canine and feline fibrosarcomas. *Journal of the American Animal Hospital Association.*, 31 (5): 439–47.
26. Shamim, Sumbul,, Ahmed, S., Waseemuddin., Azhar, Iqbal (2004) Antifungal activity of *Allium*, *Aloe*, and *Solanum* species. *Pharmaceutical Biology.*, 42 (7): 491–498.
27. Acharya SN, Thomas JE and Basu SK 2008. Fenugreek, an alternative crop for semiarid regions of North America. *Crop Science* 48, 841–853.
28. Basu SK 2006. Seed production technology for fenugreek (*Trigonella foenum-graecum* L.) in the Canadian prairies. MSc, University of Lethbridge, Lethbridge, Alberta, Canada.
29. Mir Z, Mir PS, Acharya SN, Zaman MS, Taylor WG, Mears GJ, McAllister TA and Goonewardene LA 1998. Comparison of alfalfa and fenugreek (*Trigonella foenum-graecum*) silage supplemented with barely grain on performance of growing steers. *Canadian Journal of Animal Science* 78, 343–349.
30. Montgomery J 2009. The potential of fenugreek (*Trigonella foenum-graecum*) as a forage for dairy herds in central Alberta. MSc, University of Alberta, AB, Canada.
31. Srichamroen A 2007. Hypoglycemic and hypolipidemic effects of galactomanan from fenugreek (*Trigonella foenum-graecum* L.) grown in Canada. PhD, University of Alberta, Edmonton, AB, Canada.





32. Moyer JR, Acharya SN, Mir Z and Doram RC 2003. Weed management in irrigated fenugreek grown for forage in rotation with other annual crops. *Canadian Journal of Plant Science* 83, 181–188.
33. Petropoulos GA 2002. Fenugreek – the genus *Trigonella*. Taylor and Francis, London, UK.
34. Ngugi CC, Oyoo-Okoth E, Mugo-Bundi J, Orina PS, Chemoiwa EJ, Aloo PA. Effects of dietary administration of stinging nettle (*Urtica dioica*) on the growth performance, biochemical, hematological and immunological parameters in juvenile and adult *Victoria Labeo* (*Labeo victorianus*) challenged with *Aeromonas hydrophila*. *Fish & shellfish immunology*. 2015; 44: 533-541.
35. Lans C, Turner N. Organic parasite control for poultry and rabbits in British Columbia, Canada. *Journal of ethnobiology and ethnomedicine*. 2011; 7: 21.
36. Ghasemi HA, Taherpour K, Hajkhodadadi I, Akhavan-Salamat H. Comparative effects of nettle (*Urtica dioica*) and commercial feed additives on productive performance and blood lipid profile of broiler chickens. *J Anim Sci Adv*. 2014; 4: 633-640.
37. Di Virgilio N, Papazoglou EG, Jankauskiene Z, Di Lonardo S, Praczyk M, Wielgusz K. The potential of stinging nettle (*Urtica dioica* L.) as a crop with multiple uses. *Industrial Crops and Products*. 2015; 68: 42-49.
38. Bekele AM, Beyan M, Kefyalew Berihun B. The Effect of Feeding Stinging Nettle (*Urtica Simensis* S.) Leaf Meal on Feed Intake, Growth Performance and Carcass Characteristics of Hubbard Broiler Chickens. *Global Journal of Science Frontier Research*. 2015; 15.
39. Rutto LK, Xu Y, Ramirez E, Brandt M. Mineral properties and dietary value of raw and processed stinging nettle (*Urtica dioica* L.). *International journal of food science*. 2013.
40. Kukrić ZZ, Topalić-Trivunović LN, Kukavica BM, Matoš SB, Pavičić SS, Boroja MM, et al. Characterization of antioxidant and antimicrobial activities of nettle leaves (*Urtica dioica* L.). *Acta Periodica Technologica*. 2012; 257- 272.
41. Erdogrul ÖT. Antibacterial activities of some plant extracts used in folk medicine. *Pharmaceutical Biology*. 2002; 40: 269-273.
42. Safamehr A, Fallah F, Nobakht A. Growth performance and biochemical parameters of broiler chickens on diets consist of chicory (*Cichorium intybus*) and nettle (*Urtica dioica*) with or without multi-enzyme. *Iranian Journal of Applied Animal Science*. 2013; 3:131-137.
43. Loetscher Y, Kreuzer M, Messikommer R. Oxidative stability of the meat of broilers supplemented with rosemary leaves, rosehip fruits, chokeberry pomace,



- and entire nettle, and effects on performance and meat quality. *Poultry science*. 2013; 92: 2938-2948.
44. Grata K, Nabrdalik M. Assessment of the antifungal properties of nettle extracts against *Fusarium proliferatum*. *Proceedings of ECOpole*. 2015; 9: 459-464.
45. Duda-Chodak A, Tarko T, Gajny J, Tuszyński T. Effect of different herbs on *Alicyclobacillus acidoterrestris* cultures. *Food Science and Technology*. 2009; 12: 19.
46. Stanojević LP, Stanković MZ, Cvetković DJ, Cakić MD, Ilić DP, Nikolić VD, et al. The effect of extraction techniques on yield, extraction kinetics, and antioxidant activity of aqueous-methanolic extracts from nettle (*Urtica dioica* L.) leaves. *Separation Science and Technology*. 2016; 51: 1817-1829.
47. Nissen, L. R., Byrne, D. V., Bertelsen, G., & Skibsted, L. H. (2004). The antioxidative activity of plant extracts in cooked pork patties as evaluated by descriptive sensory profiling and chemical analysis. *Meat Science*, 68, 485–495.
48. Solomakos, N., Govaris, A., Koidis, P., & Botsoglou, N. (2008). The antimicrobial effect of thyme essential oil, nisin, and their combination against *Listeria monocytogenes* in minced beef during refrigerated storage. *Food Microbiology*, 25, 120–127.
49. Camo, J., Beltrán, J. A., & Roncalés, P. (2008). Extension of the display life of lamb with an antioxidant active packaging. *Meat Science*, 80, 1086–1091.
50. Djenane, D., Sánchez-Escalante, A., Beltrán, J. A., & Roncalés, P. (2003). Extension of the shelf life of beef stakes packaged in a modified atmosphere by treatment with rosemary and display under UV-free lighting. *Meat Science*, 64, 417–426.
51. Janz, J. A. M., Morel, P. C. H., Wilkinson, B. H. P., & Purchas, R. H. (2007). Preliminary investigation of the effects of low-level dietary inclusion of fragrant essential oils and oleoresins on pig performance and pork quality. *Meat Science*, 75, 360–365.
52. Moñino, M. I., Martínez, C., Sotomayor, J. A., Lafuente, A., & Jordán, M. J. (2008). Polyphenolic transmission to Segureño lamb meat from ewes dietary supplemented with the distillate from rosemary (*Rosmarinus officinalis*) leaves. *Journal of Agricultural and Food Chemistry*, 56, 3363–3367.
53. Nieto, G., Díaz, P., Bañón, S., & Garrido, M. D. (2010). Dietary administration of ewes diets with a distillate from rosemary leaves (*Rosmarinus officinalis* L): influence on lamb meat quality. *Meat Science*, 84, 23–29.



54. Rota, M. C., Herrera, A., Martínez, R. M., Sotomayor, Jose. A., & Jordán, M. J. (2008). Antimicrobial activity and chemical composition of *Thymus vulgaris*, *Thymus zygis* and *Thymus hyemalis* essential oil. *Food Control*, 19, 681–687.
55. Kähkönen, M. P., Hopia, A. I., Heikkii, J. V., Rauha, J. P., Pihlaja, K., Kujala, T. S., et al. (1999). Antioxidant activity of plant extracts containing phenolics compounds. *Journal of Agricultural and Food Chemistry*, 47, 3954–3962.
56. Lauzurica, S., De la Fuente, J., Díaz, M. T., Álvarez, I., Pérez, C., & Cañeque, V. (2005). Effect of dietary supplementation of vitamin E on characteristics of lamb meat packed under modified atmosphere. *Meat Science*, 70, 639–646.
57. Mamta K. *Ocimum Sanctum*, as Growth Promoter in Poultry. *Dairy and Vet Sci J*. 2017; 4(5): 555647.
1. Hasan MN, Mostofa M, Sorwar MG, Hasan MT, Das K, Hossain DMN (2016) Effects of tulsi leaf extract on body weight gain in broiler production. *Bangl J Vet Med* 14(1): 21-25.
58. Biswas AK, Rahman MM, Hassan MZ, Sultana S, Rahman MM, et al. (2017) Effect of Tulsi (*Ocimum sanctum*) leaves extract as a growth promoter in broiler production. *Asian J Med Biol Res* 3(2): 226-232.
59. Alom F, Mostofa M, Alam MN, Sorwar MG, Uddin J, et al. (2015) Effects of indigenous medicinal plant tulsi (*Ocimum sanctum*) leaves extract as growth promoter in broiler. *Res Agric livest Fish* 2(1): 97-102.
60. Singh A, Doley P, Gogoi S, Neeraj (2014) Effect of dietary tulsi (*Ocimum sanctum*) leaves powder on muscle growth of broiler chicks. *Int J Biol&Pharmaceu Res* 5(1): 1-3.
61. Mode SG, Funde ST, Waghmare SP, Kolte AY (2009) Effect of herbal immune modulator on body weight gain in immunosuppressed broiler birds. *Vet World* 2(7): 269-270.
62. Gupta G, Charan S (2007) Exploring the potentials of *Ocimum sanctum* (Shyama tulsi) as a feed supplement for its growth promoter activity in broiler chickens. *Indian J Poult Sci* 42(2): 140-143.
63. Kumara M (2012) Clinico-pathological and Immunological studies in *Escherichia coli* infected broiler chicks fed on *Ocimum sanctum* leaf supplemented feed. M.V.Sc. Thesis. Lala Lajpat Rai University of Veterinary and Animal Sciences, Hisar, Haryana, India, pp.46.